

## UIC Chemistry RRC Building

### Logging into the computer and starting the NMR software TOPSPIN 1.3 on Linux CentOS

1. In the login window

login:

password:

**BOLD** lettering is typed on topspin command line or are keys pressed on board to right.

Additional information regarding topspin is on the Desktop in folder 'TOP\_processing\_a4.pdf' or 'Topspin1p3\_users\_guide.pdf'. Focus mainly on the processing, such as integration and phasing, if you get stuck.

### Changing the samples and shimming

- have the TA help you with these steps

- it is critical not to drop a sample into the magnet without hearing the air, as the sample will free fall down and break in the probe.

On the bsms board to right: a) **Spin-Off** b) **Lock-Off** c) **Lift-On**.

Sample should be swapped out, CLEANED with kim wipe, and set to probe depth using the sample gauger.

-make sure you hear the air is on before placing the sample into the magnet.

Insert sample: a) **Lift-Off** b) **Spin-On**.

**rsh shims.bbo** This will read in standard shim files.

**lock** and enter your solvent.

**lockdisp** Maximize lock (Shim) on bsms board below:

### Shim

**Z1** and use wheel to maximize lock signal. Then **Z2** and do the same. Repeat 2 times.

**STDBY** key to put the bsmsboard into standby mode.

### Proton NMR data acquisition

**edc** - Add the name as you like eg) sample\_1. This creates folder of your experiment.

**rpar h1.bbo all** – reads in 1D 1H experiment.

**ii** - this is used to initiate the interface or reset communications to the nmr.

**rga** - sets the receiver gain automatically and takes several seconds to do so.

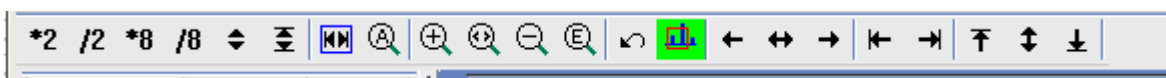
**zg** - starts experiment and overwrites current data). In pop up cl. **OK** to overwrite.

### Processing:

**efp** (when experiment finishes we fourier transform and process the data).

**apk** (autophase the spectrum, all peaks should be pointing up now). Or try manual phasing :

Adjust scaling etc with:



or

Click one of the following buttons:

**\*2** Increase the intensity by a factor of 2 [**\*2**].

**\*8** Increase the intensity by a factor of 8 [**\*8**].

**/2** Decrease the intensity by a factor of 2 [**/2**].

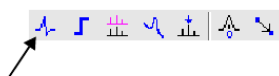
**/8** Decrease the intensity by a factor of 8 [**/8**].

**↕** Reset the intensity [**.vr**].

## Phasing

### How to Switch to Phase Correction Mode

Click the indicated button in the upper toolbar:



Click-hold the button **P** and move the mouse until the reference peak is exactly in absorption mode.

Click the button **↩** to save and execute the phase correction and return.

### Integration/peak picking:

**abs** – baseline correction

**int** – auto integrate; use *auto-find regions*

*in the integration mode (above), right click over integral region to calibrate the number of protons.*

**pps** – autopick peaks

**export** – save as *whatevername.jpg* and so on. Ok to create directory if you like.

\*Chapter 11.2 of 'Topspin1p3\_users\_guide.pdf' has much more detail regarding processing if needed.

### Acquisition of <sup>13</sup>C NMR spectrum

Create 2nd exptl data set:

**edc** and edit EXPNO entry to 2.

Note it is common practice to use the experiment # entry as:

**1:** 1D 1H

**2:** 1D 13C

**3:** Dept135 and so on. just be sure to keep track in your notes.

\* you can also change quickly between these expts once created by: **re 1** or **re 2** etc...

**edc** – create expt 2

**rpar c13.bbo all**

We use very similar steps as in the 1H expt.

**ii**

**zg** (experiment takes 5 minutes).

\*If signal to noise is still too low you can increase the number of scans **ns** (eg 2X-4X's more) and type **go** to continue signal averaging onto the previous FID.

**efp** - process the data

**apk** - autophase. Now adjust peaks intensities so they fit to screen. use the \*2 or /2 buttons.

Or phase manually again as well.

**setti** -include appropriate title

**pps** – autopick peaks

**export** – save as *whatevername.jpg* and so on. Ok to create directory if you like.

**Additional experiments can be collected in a very similar fashion (please see the end of this document). All experiments are easily setup using the 'rpar' command.**

Acquisition of DEPT-135 13C NMR spectrum (CH3/CH up; CH2 Down)

**edc** – create expt 3

**rpar dept135.bbo all**

**ii**

**zg** (experiment takes 5 minutes).

Manually phase from above. Note some peaks are supposed to point down if present.

**setti** -include appropriate title

**pps** – autopick peaks

**export** – save as *whatevername.jpg* and so on. Ok to create directory if you like.

Acquisition of DEPT-90 13C NMR spectrum (CH only up)

**edc** – create expt 4

**rpar dept90.bbo all**

**ii**

**zg** (experiment takes 5 minutes).

Manually phase from above.

**setti** -include appropriate title

**pps** – autopick peaks

**export** – save as *whatevername.jpg* and so on. Ok to create directory if you like.

Acquisition of DEPT-45 13C NMR spectrum (CH3/CH2/CH all up)

**edc** – create expt 5

**rpar dept45.bbo all**

**ii**

**zg**

Manually phase from above. All peaks point up.

**setti** -include appropriate title

**pps** – autopick peaks

**export** – save as *whatevername.jpg* and so on. Ok to create directory if you like.  
Export your data..

#### 2D HMQC (1H/13C) Correlation via 1 Bond (direct attachment)

**edc** – create expt 6  
**rpar hmqc.bbo all**  
**ii**  
**1 td 128**  
**zg**

*2D processing (for cosy and hmqc above)*  
**xfb** – does 2d fourier transform and phase correction.

\*scaling of intensities/contours down just like with the 1Ds.  
\*use LMB and box in area of zoom if you like.  
**export** – save as *whatevername.jpg*

#### 2D COSY (1H/1H) Correlation

**edc** – create expt 7  
**rpar cosy.bbo all**  
**ii**  
**zg**

*2D processing (for cosy and hmqc above)*  
**xfb** – does 2d fourier transform and phase correction.

\*scaling of intensities/contours down just like with the 1Ds.  
\*use LMB and box in area of zoom if you like.  
**export** – save as *whatevername.jpg*

#### When finished email yourself the data

\*data .jpgs are stored on desktop

#### Finishing up with TOPSPIN and logging off the computer

36. Remove your sample and replace it with the CDCl<sub>3</sub> standard following steps 3, 4 and 5 above again.

37. Type **exit** to leave the NMR program.

38. logout icon (> type arrow) is on top bar.

39. **rpar** (will give a list of all the expts available. You should focus only ones in **lower case** as expts in ALL CAPS are from standard bruker files). These however can be used too with **prosol** setup methods and please let me know if you'd like to try.